

Original Research Article

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## Mass Multiplication of Entomopathogenic Nematodes on *in vitro* Solid Media

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### ABSTRACT

#### Keywords

*H. indica*, Wout's medium, Entomopathogenic nematodes, *S. abbasi*

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In this study, two Haryana local isolates of entomopathogenic nematodes (EPNs) namely *Steinernema abbasi* and *Heterorhabdus indica* were mass multiplied on eight culture media comprising of Wout's medium, Modified Wout's medium I, Modified Wout's medium II, Modified Wout's medium III, Egg yolk medium I, Egg yolk medium II, Modified egg yolk medium and Modified dog biscuit medium and harvested at two different time intervals *i.e.* at 30 days and 60 days interval. Along with this, studies were also conducted to know the effect of temperature *i.e.* at 15°C, 20°C and 30 °C on the infective juvenile (IJ) yields. Of the eight media tested, *S. abbasi* and *H. indica* both multiplied maximum on modified dog biscuit medium. Wout's medium was second best in IJs yield. Yield was maximum at 30 days and reduced drastically at 60 days. *S. abbasi* yielded maximum at 25 °C whereas *H. indica* multiplied maximum at 30 °C. None of the species multiplied at 15 °C.

### Introduction

The nematodes belonging to the genus *Heterorhabditis* and *Steinernema* are by fame and name noted to be successful bio agents as they are used to control insect pests. These EPNs are symbiotically associated with bacteria of the genera *Photorhabdus* and *Xenorhabdus* that belongs to Enterobacteriaceae. EPNs have broad host range and exempted from the registration requirements in several countries (Kaya and Gaugler, 1993). The most important attribute

is that they can be applied in conjugation with fertilizers, soil amendments and insecticides as upto 2-5 hours infective juveniles (IJs) are tolerant to their exposure (Rovesti *et al.*, 1988; Rovesti and Desco, 1990). Tank mixed EPNs and pesticide combinations offer the cost effective alternative in Integrated Pest Management (IPM) as EPNs are safe and environmental friendly. For these aspects EPNs are required in bulk to apply in field level and they can be multiplied in laboratory or by commercial companies *in vitro* solid and liquid media.

Based on the attractive attributes of entomopathogenic nematodes, the identified indigenous species are to be cultured *in vivo* or *in vitro* for large scale commercial production as well as for laboratory experimentation and field testing (Shapiro-Ilan *et al.*, 2002). Since forties, the technology of *in vitro* mass rearing of EPNs has been developed (Glaser, 1940). At times EPNs are produced by variety of means by insect infection or on artificial media (axenic or monoxenic culture) in solid and liquid phase of fermentation. They are produced commercially by more than 10 countries in Asia, Europe and North America and to date more than 12 different species have reached a commercial development: *H. bacteriophora* Poinar 1976, *H. indica* Poinar *et al.*, 1992, *H. marelatus* Liu and Berry 1966, *H. megidis* Poinar *et al.*, 1987, *H. zealandica* Poinar 1990, *S. carpocapsae*, *S. feltiae* (Filipjev 1934) Wout's *et al.*, 1982, *S. glaseri*, *S. kushidai* Mamiya 1988, *S. kraussei* (Steiner, 1923) Travassos 1927, *S. longicaudatum* Shen and Wang 1992, *S. riobrave* and *S. scapterisci* Ngunyun and Smart 1990 (Lacey *et al.*, 2001).

For a biocontrol agent to be successful it should be amenable for mass production on a large scale, the ready availability of the organism in required quantity and at competitive cost makes them acceptable among entrepreneurs and farmers. Over the years many isolates of EPNs have been collected from Haryana soils. Though Haryana is a small state it represents diverse agro-ecological zones. The north eastern zone comprises of Shivalik hilly region and alluvial plains and the western zone consists mainly of alluvial plain. The major pests are rice root weevil, leaf folder, brown plant hopper, stem borer, sugarcane pyrrilla, cotton boll worms, mustard aphids, cabbage diamond back moths, *etc.* Hence for the successful control of these pests the local isolates have to be collected, identified up to species level and their mass multiplication methods are to be standardized

to use them in field against insect pests.

## Materials and Methods

The culture of indigenous isolates of *S. abbasi* and *H. indica* were obtained from the Department of Nematology, CCSHAU, Hisar, Haryana. The cultures were multiplied on the last instar larvae of *G. mellonella*. Conical flasks, Petri plates, beakers, watch glasses were placed in hot air oven at 180 °C for 1 h before use. Forceps and needles were washed with absolute ethanol before use.

## Composition of the eight media used (Hussaini *et al.*, 2002)

- a. Wout's medium: nutrient broth- 0.88 g, yeast extract- 0.32 g, soy flour- 14.4 g, groundnut oil-10.4 g and distilled water- 60 ml.
- b. Modified Wout's medium I: nutrient broth- 0.88 g, yeast extract- 0.32 g, corn flour- 14.4 g, groundnut oil-10.4 g and distilled water- 60 ml
- c. Modified Wout's medium II: nutrient broth- 0.88 g, yeast extract- 0.32 g, soy flour- 14.4 g, sesamum oil-10.4 g and distilled water- 60 ml
- d. Modified Wout's medium III: nutrient broth- 0.88 g, yeast extract- 0.32 g, corn flour- 14.4 g, sesamum oil-10.4 g and distilled water- 60 ml
- e. Egg yolk medium I: egg yolk- 7.0 g, yeast extract- 2.0 g, sodium chloride- 0.8 g, groundnut oil- 15.0 g and distilled water- 60 ml
- f. Egg yolk medium II: egg yolk – 10.0 g, yeast extract- 5.0 g, sodium chloride- 0.8 g, groundnut oil- 12.0 g and distilled water- 60 ml
- g. Modified egg yolk medium: egg yolk- 7.0 g, soy flour- 20.0 g, yeast extract- 2.0 g, sodium chloride- 0.8 g, groundnut oil- 15.0 g and distilled water- 60 ml
- h. Modified dog biscuit medium: dog biscuit

20.0 g, peptone- 0.5 g, yeast extract 1.0 g, beef extract 5.0 g, groundnut oil- 7.0 g and distilled water- 100 ml

### **Preparation of media for the multiplication of EPNs**

In each medium the ingredients were weighed according to the requirement and mixed in distilled water. The medium was gently heated and stirred uniformly.

In one basin sponge material (polyether polyurethane foam) which was cut into small pieces of  $1 \times 1 \text{ cm}^3$  was taken and washed thoroughly with distilled water 2 to 3 times. These were dried at room temperature. The medium was poured into the basin containing the sponge and mixed thoroughly, so that it was coated uniformly onto foam pieces (1.5 g foam chips, 8 to 9 g medium w/w).

The conical flasks (250 ml capacity) were filled with 8 to 10 media coated sponges. The mouth of the flask was cleaned to remove the adhered medium and plugged tightly with non-absorbent cotton. These flasks were autoclaved at  $121.6^\circ\text{C}$  for 20 to 30 minutes. After autoclaving, the flasks were allowed to cool before use.

### **Preparation of nutrient agar and nutrient broth**

For the preparation of nutrient agar, the ingredients (peptone- 5.0 g, NaCl- 5.0 g, beef extract- 3.0 g, yeast extract- 2 g, agar- 20 g) were weighed and mixed in 1 litre distilled water, gently heated by stirring uniformly.

The medium was poured in conical flasks (250 ml capacity) plugged with cotton and autoclaved at  $121.6^\circ\text{C}$  for 15 to 20 min. Nutrient broth also prepared as above separately in which agar was not added.

### **Isolation and preparation of symbiotic bacterial cultures**

The isolation of the symbiotic bacteria from the haemolymph of *Galleria mellonella* Linnaeus (greater wax moth) larvae was done following the method of Kaya and Stock 1997. Under a laminar flow sterile bench, late instar larvae of *G. mellonella* were placed in a Petri dish with moist filter paper and inoculated with IJs separately for each nematode species (100 IJs per larva). The cadavers so obtained were surface sterilized with 1.0 % (w/v) sodium hypochlorite solution and rinsed three times with sterilized water.

The cadaver was dipped in absolute ethanol, ignited and immediately plunged into sterile water. Using the sterile injection needles the cadaver was carefully opened from the ventral side. Then using a sterile inoculation loop, the haemolymph of the cadaver was slightly touched and transferred to the nutrient agar plates by streaking in a zig-zag manner near the burner. The Petri plates were then properly labeled, put in polythene cover and kept at  $25 \pm 1^\circ\text{C}$  for 48 h in an BOD incubator.

### **Multiplication of symbiotic bacteria**

In the laminar flow chamber, the single colonies of the bacterium in the nutrient agar were gently touched with sterile inoculating loop and transferred to the nutrient broth in flasks. The flasks inoculated with the bacterium, were wrapped with black paper, kept on the mechanical shaker and continuously stirred at room temperature for 24 to 48 h. After 48 h bacterium culture so obtained was inoculated to the media in flasks at 0.5 ml per flask, kept in dark at  $25 \pm 1^\circ\text{C}$  in a BOD incubator.

### **Surface sterilization of IJs**

The IJs were collected in 100 ml of clean sterile distilled water. The IJs were surface

sterilised by adding 1 ml 0.25% (w/v) Hyamine (methylbenzethonium chloride) to the suspension and washed with sterile water 2 to 3 times in the laminar air flow chamber. With a sterile micropipette, most of the supernatant was removed without disturbing the IJs at the bottom of the tube. Each step was started with enough IJs to compensate for some loss during the process. At the final step the surface sterilized IJs which were collected in a beaker with sterile distilled water. The suspension was adjusted so that 1 ml contained approximately 2000 IJs.

### **Inoculation of IJs into the medium and incubation of media**

The nematodes were inoculated in the flasks @ 2000 IJs per flask (250 ml) under aseptic conditions. The cotton plugged flasks were wrapped with paper to ensure dark conditions and incubated at  $25\pm 1^\circ\text{C}$  for 30 and 60 days. The flasks were checked for contamination on every alternate day. Each treatment was replicated four times. IJs of *S. abbasi* were inoculated in the flasks containing *Xenorhabdus* bacterium and IJs of *H. indica* were inoculated into flasks containing *Photorhabdus* bacterium.

### **Harvesting of IJs**

After the incubation for 30 and 60 days at  $25\pm 1^\circ\text{C}$ , the IJs were extracted from the different media by pouring out the coated sponge media on 400 mesh sieve placed in the water filled basin for 1 h. The IJs were collected from the basin and concentrated in a beaker. The nematodes were counted under the microscope by the dilution method and the final observation on yield (number of IJs) in each flask was recorded.

### **Statistical analysis**

The data were analysed statistically by using Completely Randomized Design (CRD)

following 'OP Stat' available at CCS HAU, Hisar, website.

### **Results and Discussion**

A steinernematid, *S. abbasi* and a heterorhabditid, *H. indica* were inoculated on eight different media (composition as given in materials and methods) to select the most suitable medium for multiplication. The number of IJs recorded after 30 and 60 days of inoculation are put together in Table 1. The data revealed that both the species multiplied maximum on modified modified dog biscuit medium (71,56,071 and 10,21,634 IJs per 250 ml flask of *S. abbasi* and *H. indica* (Fig. 2a, 3a), respectively). *S. abbasi* yield was significantly more than *H. indica* in all the media except Wout's medium where *H. indica* IJs recovered were more (6,99,100 IJs per 250 ml flask) compared to *S. abbasi* (58,333 IJs per 250 ml flask). In all other media, *S. abbasi* multiplied better than *H. indica*. Out of eight media tested, four viz., modified Wout's medium II, egg yolk medium I, II and modified egg yolk medium were unsuitable for multiplication, since there were no IJs recovery of the two species in these media except in egg yolk medium I where a very low yield of *S. abbasi* (2,664 IJs per 250 ml flask) was obtained.

The order of suitability of the media for the multiplication of *S. abbasi* was modified dog biscuit medium (71 lakh IJs), modified Wout's medium III (2.07 lakh IJs), modified Wout's medium I (77 thousand IJs), Wout's medium (58 thousand IJs) and egg yolk medium I (2.6 thousand IJs) whereas for *H. indica* modified dog biscuit medium (10 lakh IJs) was followed by Wout's medium (6.99 lakh IJs), modified Wout's medium III (31.7 thousand IJs) and modified Wout's medium I (15.9 thousand IJs).

The recovery of IJs was reduced after 60 days so much so that there were 36 % less IJs in *S.*

*abbasi* and 65 % less in *H. indica* from modified dog biscuit medium. In all other media too, there was 30- 70 % reduction in IJs yield in both the nematode species.

Of the eight media used, Wout’s medium and modified dog biscuit medium were selected to study the effect of temperature on IJs yield of both the nematode species. At 15°C, *S. abbasi* gave similar yield in both the media where as *H. indica* did not multiply in any of the two

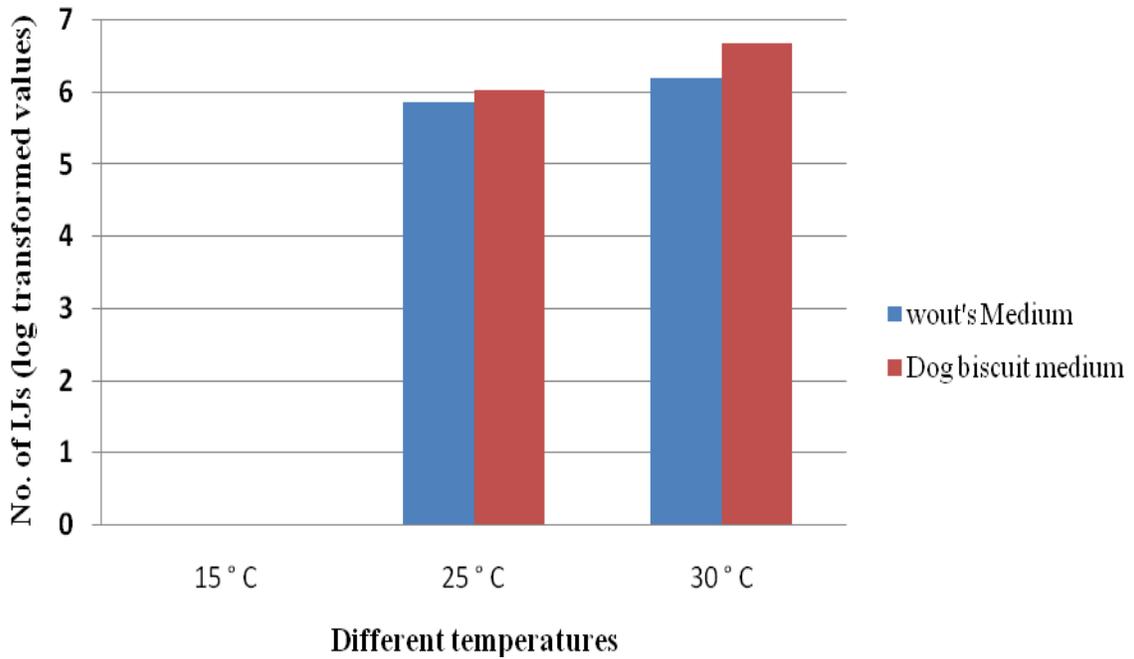
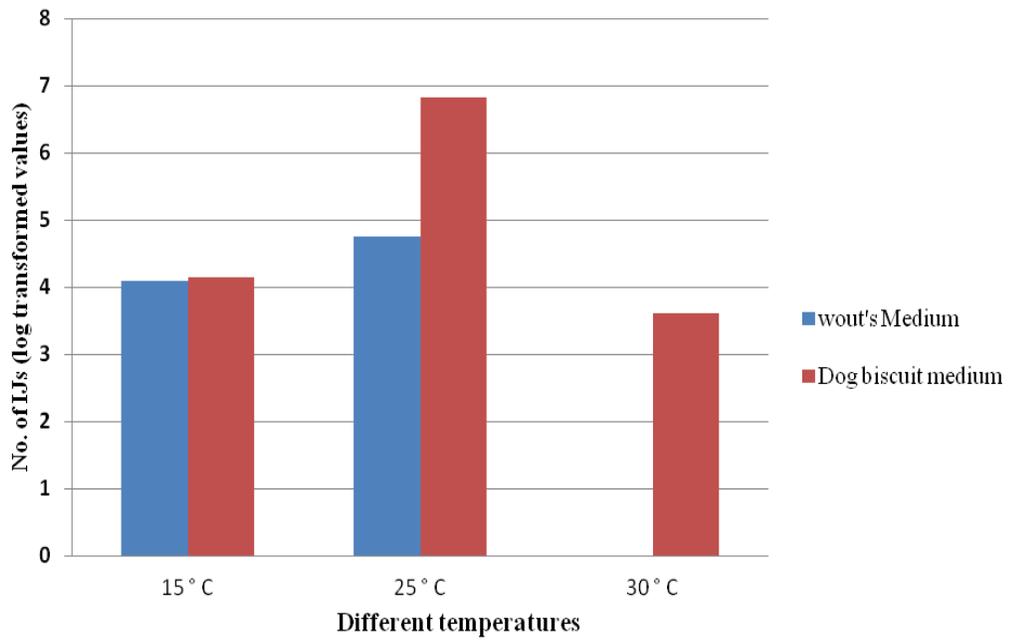
media (Fig. 1A, B, 4 and 5). *S. abbasi* multiplied maximum in modified dog biscuit medium at 25°C followed by Wout’s medium at the same temperature. *S. abbasi* did not multiplied in Wout’s medium at 30°C while in modified dog biscuit medium the yield was low compared to 25°C. *H. indica* in comparison to *S. abbasi* multiplied better in both the media at 30°C than at 25°C and more so in modified dog biscuit medium.

**Table.1** Number of infective juveniles produced on different media at two time intervals (Mean of four replications)

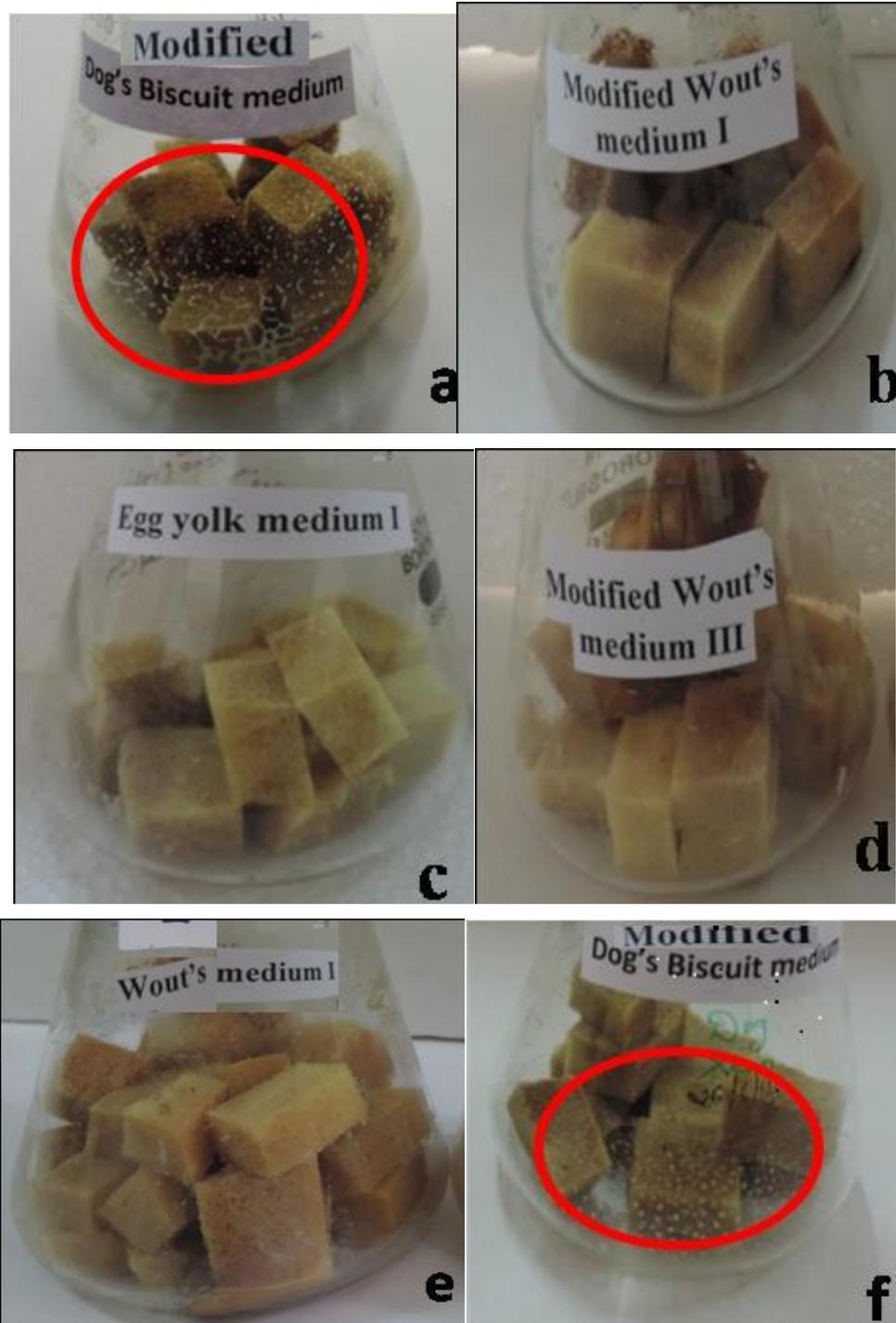
Nematode species	<i>Steinernema abbasi</i>		<i>Heterorhabditis indica</i>	
	30 DAYS	60 DAYS	30 DAYS	60 DAYS
Wout’s medium	58,333 (4.77)	10,289 (4.01)	6,99,100 (5.84)	1,29,450 (5.10)
Modified Wout’s medium I	77,389 (4.89)	24,375 (4.39)	15,949 (4.20)	7,225 (3.85)
Modified Wout’s medium II	-	-	-	-
Modified Wout’s medium III	2,07,455 (5.27)	52,408 (4.57)	31,770 (4.50)	24,330 (4.37)
Egg yolk medium I	2664 (3.41)	-	-	-
Egg yolk medium II	-	-	-	-
Modified Egg yolk medium	-	-	-	-
Modified dog biscuit medium	71,56,071 (6.85)	45,90,469 (6.66)	10,21,634 (6.00)	2,97,646 (5.47)
CD at 5%	(0.18)	(0.33)	(0.06)	(0.17)

Figures in parentheses are log transformed values

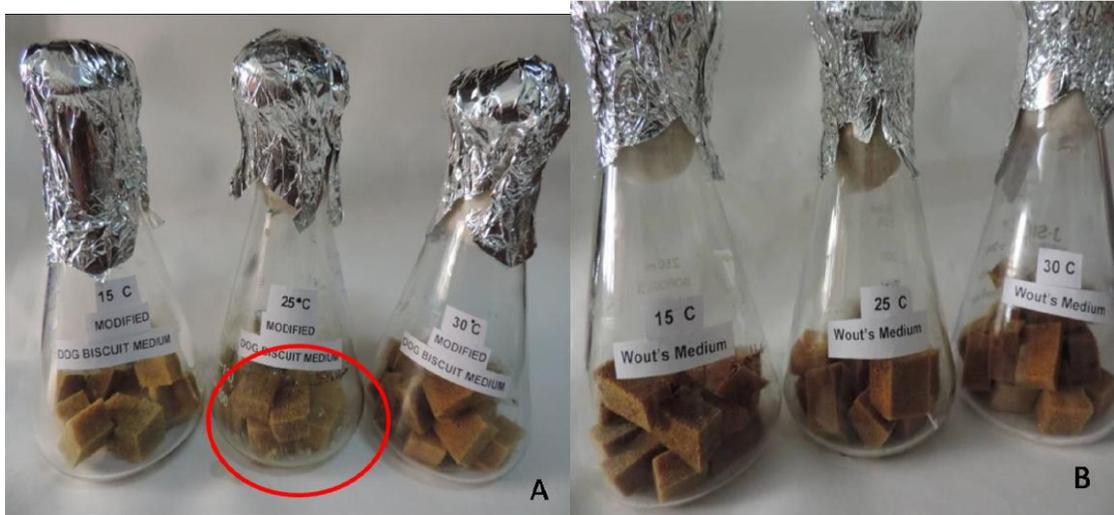
**Fig.1** Multiplication of (A) *Steinernema abbasi* & (B) *Heterorhabditis indica* on modified dog biscuit medium and Wout's medium at different temperatures. (Mean of four replications)



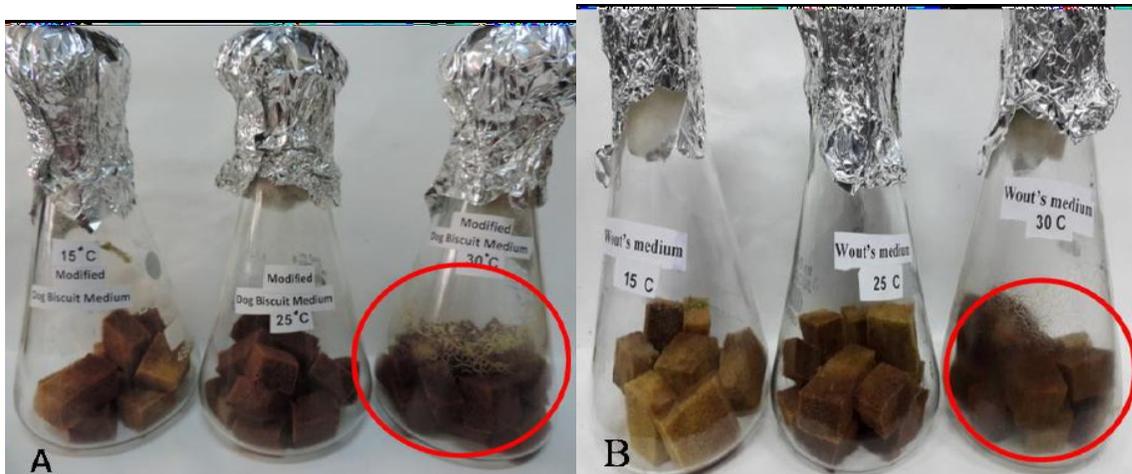
**Fig.2** Multiplication of *Steinernema abbasi* after 30 days of nematode inoculation; a) Modified dog biscuit medium, b) Modified Wout's medium I, c) Egg yolk medium I, d) Modified Wout's medium III e) Multiplication of *Steinernema abbasi* on modified dog biscuit medium after 60 days of inoculation



**Fig.3** Multiplication of *Heterorhabditis indica* after 30 days of nematode inoculation; a) Modified dog biscuit medium, b) Modified Wout's medium I, c) Modified Wout's medium III, d) Wout's medium e) Multiplication of *Heterorhabditis indica* after 60 days of nematode inoculation on modified dog biscuit medium



**Fig.5** Multiplication of *Heterorhabditis indica* on A) Modified dog biscuit medium and B) Wout's media at three different temperatures



Entomopathogenic nematode's potential as efficient biocontrol agents can only be assessed by field applications for which large number of infective juveniles (IJs) are required. For small scale experimentation, IJs can be multiplied on natural host or alternate host, *G. melonella* being the most preferred for *in vivo* culturing of the majority of the

species. This method is not cost effective and involves a lot of labor. So, efforts in the direction to culture these *in vitro* were initiated by Glaser as early as 1940 on a solid medium axenically. Soon, it was established that inoculation of natural symbiotic bacterium to the medium before nematode inoculation increased the yields considerably

(Poinar and Thomas, 1966). Also the primary form of bacterial symbiont is better suited for EPNs (particularly *Heterorhabditis*) than secondary form. Wout's 1981 and Bedding 1981 used plant protein origin and animal protein origin media for culturing EPNs *in vitro*. To increase the surface area polyether polyurethane sponge was used. Plant protein origin media are preferred since animal protein origin media smell awfully.

Keeping all the conditions in view, five different media of plant protein origin *viz.* Wout's, Wout's I, Wout's II, Wout's III and modified dog biscuit (purely of vegetative origin) media and those of animal protein origin (Egg yolk I, Egg yolk II and Egg yolk III) were used to culture indigenous isolates of *S. abbasi* and *H. indica* to select the most suitable medium. In our study animal protein media (all Egg yolk media) did not yield any progeny. Modified dog biscuit medium (consisting of wheat flour, oats, corn, rice and vegetables supplemented with herbal liver tonic) yielded maximum yielded maximum in both the species, *S. abbasi* and *H. indica*. The yield was higher in *Steinernema* compared to *Heterorhabditis* though no particular reason could be worked out yet the possibility of changing primary form of bacterium to secondary form can be one of the reasons. Dog biscuits were successfully as early as 1965 by House *et al.*, in Canada to mass multiply *S. carpocapsae* (DD 136 strain). Similar results were also obtained by Hara *et al.*, (1981) and Kondo and Ishibashi in 1985 and 1991. In all these studies dog biscuit or dog food medium were of animal protein origin whereas in the present study dog biscuits were of plant protein origin.

Hussaini *et al.*, (2002) reported that *H. indica* (PDBC EN6.7) multiplied well on Wout's medium and modified Wout's medium containing soy flour and groundnut oil in various proportions. However in our studies

*H. indica* yielded more in medium with soy flour and groundnut oil whereas *S. abbasi* preferred corn flour and sesamum oil medium. Ogura and Haraguchi (1993) reported that in general plant protein origin media supported low multiplication compared to animal protein origin which is in contrast to our observations where plant protein based media yielded more. Also the report of Pervez and Ali (2010) that *S. masoodi*, *S. mushtaqi* and *S. seemae* multiply more on egg yolk medium is not consumerate with our studies where both the species did not multiply on this medium or its modifications thereof. Our study also not gained support from Prabhu *et al.*, (2006) who reported more multiplication on animal protein medium than plant protein medium. The variation can be due to different species/ isolate used since EPN species behave differently in different environments. Hence, there is a need to standardize the medium and the culture conditions for every new isolate to obtain maximum yield. The cultures yielded maximum after 30 days which decreased drastically at 60 days. The plausible reason for this can be the depletion of the food material and/or accumulation of toxic metabolites. Hence, the cultures are to be subcultured every month to maintain their viability and growth potential.

The entomopathogenic nematodes isolated from temperate areas are adapted to low temperature conditions whereas those isolated from tropical or sub tropical areas can tolerate warmer conditions. There is a variation in this temperature tolerance in different species of two genera and a set of temperature conditions are required for their growth and multiplication. In the present investigations, both the species were isolated from the Haryana soils even then there was variation in their temperature requirement. *S. abbasi* yielded more at 25°C whereas *H. indica* multiplied maximum at 30°C. In *H. indica* IJs, second stage cuticle is retained in most of the

specimens which perhaps renders it more tolerant to environmental conditions. Similar results were obtained by Yamanaka *et al.*, (2000) who reported 25°C as temperature suitable for multiplication of *S. glaseri* which did not survive at and above 30°C. Han *et al.*, (1993) too observed that normal temperature range for IJ production of *S. carpocapsae* was 19-27°C and below 15°C and above 31°C no offspring produced due to increase in mortality. *H. bacteriophora* multiplied maximum at 28±2°C (Floyd *et al.*, 2012). In Pakistan, Salma and Shahina (2012) observed that all the EPN strains tested multiplied at 32±2°C except *S. feltiae* which reproduces better at 20°C since this is a cold tolerant species.

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